# Folded Structures in Protonated Reduced Dipeptides

## VINCENT GRAND,<sup>1</sup> ANDRÉ AUBRY<sup>2</sup>, VIRGINIE DUPONT,<sup>1</sup> ANDRÉ VICHERAT<sup>1</sup> and MICHEL MARRAUD<sup>1</sup>

<sup>1</sup>Laboratory of Macromolecular Physical Chemistry, ENSIC-INPL, Nancy, France <sup>1</sup>Laboratory of Cristallography and Modelling of Mineral and Biological Materials, University of Nancy I, Vandoeuvre, France

Recevied 12 December 1995 Accepted 1 April 1996

Abstract:Reduced dipeptides with the general formula RCO-Xaa-rXbb-N<sup>+</sup>HR'R" (rXbb, reduced analogue of residue Xbb: NH-C"HR<sup>1</sup>-C<sub>r</sub>H<sub>2</sub>) are shown to adopt a folded conformation in solution and in the solid state. The protonated reduced amide bond is an active proton donor capable of interacting with a peptide carbonyl to give a strong hydrogen bond topologically equivalent to the i+2 or  $i+3 \rightarrow i$  interaction. The resulting conformation is similar to the  $\gamma$ - or  $\beta$ -turn structure found in peptides and proteins.

Keywords: conformational analysis; crystal structure; folded structures; pseudopeptides; reduced peptides

Because of its resistance against enzymic degradation, the reduced amide bond (CH<sub>2</sub>—NH) has been largely used for the design of peptidase inhibitors [1– 6] and introduced in bioactive peptide analogues [6– 14]. Despite their common use, the reduced peptides have been the subject of only a limited number of conformational NMR [11, 12, 15–18] and theoretical analyses [19, 20]. The structures of some reduced protease-inhibitor complexes have been solved by Xray diffraction, but the resolution accuracy is rather low [2, 3, 5]. The crystal structures of N<sup> $\alpha$ </sup>-Z and N<sup> $\alpha$ </sup>-Boc-Pro $\psi$ [CH<sub>2</sub>—NH]Leu-Gly-NH<sub>2</sub>, derived from the oxytocin C-terminal tripeptide and containing a reduced Pro-Leu amide bond, have been reported [21].

In previous investigations we have studied two reduced analogues of the Piv-Pro-Gly-NHR' model dipeptide, containing a C-terminal reduced amide bond [22]. The reduced amide (amine) group, may be protonated at the physiological pH, and we have shown that the protonated reduced amide link is a strong proton-donating group capable of interacting with the acyl carbonyl. The resulting folded structure, resembling the  $\beta$ II-turn in peptides, has been solved by X-ray diffraction when the protonated reduced peptide is associated with the BPh<sub>4</sub><sup>--</sup> anion [22, 23].

The present study reports the conformational analysis of similar reduced dipeptides, derived from the Pro-Gly, Pro-Ala, Gly-Ala and Ala-Pro sequences, and having the general formula RCO-Xaa-rXbb-N<sup>+</sup>HR'R" (Table 1), where rXbb denotes the reduced amino acid residue NH-C<sup> $\alpha$ </sup>HR<sub>c</sub>-C<sub>r</sub>H<sub>2</sub>. In order that the intramolecular interactions could prevail over the ammonium–anion interactions, the protonated reduced dipeptides have been associated with the weakly polar BPh<sub>4</sub><sup>-</sup> or PF<sub>6</sub><sup>-</sup> anion [24, 25], and examined by FT-IR and <sup>1</sup>H-NMR in organic solution. The crystal structure of Piv-Pro-rAla-N<sup>+</sup>HMe<sub>2</sub>, BPh<sub>4</sub><sup>-</sup> has been solved by X-ray diffraction.

Abbreviations: b. broad; COSY, correlated spectroscopy; d. doublet; DCC, *N.N*-dicyclohexylcarbodiimide; DCM, dichloromethand; DMA, dimethylacetamide; DMAP, 4-dimethylamino-pyridine; DMSO, dimethylsulphoxide; FT-IR, Fourier-transform infrared spectroscopy; *J*, coupling constant, m, multiplet; NMM, *N*methylmorpholine; ONp, 4-nitrophenyl; Ph, phenyl; Piv, pivaloyl; rXaa or rXbb, reduced amino acid residue; s, singlet; TOCSY, totally correlated spectroscopy; Xaa or Xbb, amino acid residue; Z, benzyloxycarbonyl.

Address for correspondence: Dr Michel Marraud, Laboratory of Macromolecular Physical Chemistry ENSIC-LCPM, BP 451, 54001 Nancy Cedex, France. Tel (+33) 83 17 51 91; fax (+33) 83 37 99 77).

<sup>© 1996</sup> European Peptide Society and John Wiley & Sons, Ltd. CCC 1075-2617/96/060381-11

Table 1The Protonated Reduced Dipeptides Inves-tigated with Their Given Codes<sup>a</sup>

Protonated reduced dipeptide	Code	Anion
Piv-Pro-rAla-N <sup>+</sup> HMe <sub>2</sub>	PrA	BPh <sub>4</sub> <sup>-</sup>
Piv-Pro-rGly-N <sup>+</sup> HMe <sub>2</sub> <sup>b</sup>	PrG	$BPh_4^-$ or $PF_6^-$
Piv-Pro-rGly-N <sup>+</sup> H <sub>2</sub> Et <sup>b</sup>	PrG′	$BPh_4^-$
Boc-Gly-rAla-N <sup>+</sup> HMe <sub>2</sub>	GrA	BPh <sub>4</sub> -
Piv-Gly-rGly-N <sup>+</sup> H <sub>2</sub> Et <sup>c</sup>	GrG′	$BPh_4^-$
Boc-Ala-rPro-N <sup>+</sup> H <sub>2</sub> Me	ArP	BPh <sub>4</sub> <sup></sup>
Piv-rPro-N <sup>+</sup> H <sub>2</sub> iPr	rP	$PF_6^-$

<sup>a</sup>The letter r preceding the three- or one-letter code denotes the reduced anlogue (NH-C<sup>x</sup>HR<sub>r</sub>CH<sub>2</sub>) of the peptide residue. The rAla and rPro reduced residues have the same absolute S-configuration as the cognate L-Ala and L-Pro residues.

<sup>b</sup>Reference [22].

<sup>c</sup>Reference [23].

#### **Crystal structures**

The bond lengths and bond angles of the protonated reduced amide group  $C^{\alpha}-C_{r}-N^{+}-C$  in **PrA** and **GrG'** are indicated in Figure 1. Due to the absence of electronic conjugation, the  $C_{r}-N^{+}$  bond is significantly longer, and both  $C^{\alpha}-C_{r}-N^{+}$  and  $C_{r}-N^{+}-C$  angles significantly smaller than their homologues in peptides [37]. However, the  $C^{\alpha}\cdots C$  distance of about 3.8 Å between the transoid carbons is unchanged with reference to peptides. The N<sup>+</sup>-H site in **PrA** is strongly intramolecularly hydrogen bonded to the Piv-carbonyl, with a short N<sup>+</sup> $\cdots$ O distance of 2.82 Å, in a similar way to the  $i+3 \rightarrow i$  interaction typical of the  $\beta$ -turn structure in peptides [38]. The same interaction is also present in the crystal structures of **PrG**, **PrG'** and **GrG'** [22, 23].

All the crystallized protonated reduced dipeptides adopt a folded conformation which, according to the Pro- $\psi_1$  angle (Table 2), resembles the  $\beta$ II-turn in peptides [38]. Actually, these reduced dipeptides assume two conformations, noted  $\beta$ II<sup>+</sup> and  $\beta$ II<sup>-</sup> (Figure 2), differing in the rotational states of the



Figure 1 Dimensions of the protonated reduced amide link in **PrA** (a) and **GrG**' (b) [23].

N–C<sup> $\alpha$ </sup> and C<sup> $\alpha$ </sup>–C<sub>*r*</sub>H<sub>2</sub> bonds (' $\phi_2$ ', ' $\psi_2$ '  $\approx$  50°,50° or 90°,-60°). Conformation  $\beta$ II<sup>+</sup> places the (rAla)C<sup> $\beta$ </sup>H<sub>3</sub> group of **PrA** in a pseudo-equatorial orientation with respect to the 10-membered ring (Fig. 3(a)). Both conformations  $\beta$ II<sup>+</sup> and  $\beta$ II<sup>-</sup> are present in the crystal structure of **PrG** [22].

There is no intermolecular hydrogen bond in the crystal structure of **PrA**. The (rAla)N-H group is in contact with the BPh<sub>4</sub><sup>-</sup> anion as illustrated in Figure 3 with three short N····C distances of 3.38–3.46 Å. In Figure 3 we also note the stacking of the Pro pyrrolidine cycle with one BPh<sub>4</sub><sup>-</sup> aromatic ring.



Figure 2 Stereoviews of the folded crystal molecular conformation of **PrA** (a,  $\beta II^+$ -type) and **PrG** (b,  $\beta II^-$ -type [22]) stabilized by an N<sup>+</sup>-H···O=C interaction closing a 10-membered cycle.



Figure 3 Stereoviews showing the relative disposition of the  $BPh_4^-$  anion (thin line) and the protonated reduced dipeptide **PrA** (heavy line) in the crystal.

	PrA <sup>a</sup>	Рг	G <sup>b</sup>	Pi	rG′	<b>GrG</b> ′ <sup>c</sup>
Conformation <sup>d</sup>	$\beta$ II $^+$	etaII+	βII <sup>-</sup>	βII <sup></sup>	$\beta$ II $^-$	βII <sup>-</sup>
ω <sub>0</sub>	175.6(5)	178	178	-178	180	-176
Pro/Gly						
$\phi_1$	-52.4(8)	-53	-53	~58	-48	-53
$\psi_1$	140.5(5)	136	136	145	141	140
$\omega_1$	-178.2(6)	-171	-171	-173	-177	-167
rAla/rGly <sup>e</sup>						
'φ <sub>2</sub> '	52.4(8)	41	97	95	99	92
ʻψ <sub>2</sub> `	41.5(8)	53	-63	~61	-63	-63
'ω <sub>2</sub> '	-171.7(6)	-170	167	-176	-177	-178
	62.2(8)	60	-72			

Table 2Backbone Torsional Angles (degrees) in the Molecular Structures of Crystallized Protonated ReducedDipeptides

<sup>a</sup>Present work.

<sup>b</sup>Two independent molecules per asymmetric unit [22].

<sup>c</sup>Reference [23].

<sup>d</sup>This folded conformation is denoted with reference to the cognate  $\beta$ -turn type, indicating in superscript the sign of angle ' $\psi_2$ '. <sup>c</sup>The torsional angles in the reduced rAla/rGly residue are defined as follows: ' $\phi' = C - N - C^{\alpha} - C_r$ , ' $\psi' = N - C^{\alpha} - C_r - N^+$ ; ' $\omega' = C^{\alpha} - C_r$  $-N^+ - C$ 

#### Conformations in solution

The conformational preferences of such small protonated reduced dipeptides in organic solution is essentially governed by the formation of intramolecular hydrogen bonds, which can be revealed by the shift to lower frequencies of both the N--H and C=O stretching IR absorptions.

## PrA and PrG Reduced Peptides

**PrG** and **PrA**, associated with the  $BPh_4^-$  anion, exhibit in DCM very similar IR data (Table 3): (1) the low (Piv)C=O absorption near 1585 cm<sup>-1</sup>, superimposed on the weak and sharp BPh<sub>4</sub><sup>-</sup> aromatic contribution at 1580  $\text{cm}^{-1}$ , denotes the participation of this vibrator in a strong hydrogen bond; (2) the (Pro)C=O absorption at 1687  $\text{cm}^{-1}$  is typical of a free vibrator; (3) the absence of any absorption at about 3100 cm<sup>-1</sup>, expected for the  $N^+$ -H $\cdots$ BPh<sub>4</sub><sup>-</sup> ionic pair, and the presence of a broad and multicomponent absorption band centered at 2755  $cm^{-1}$ are in favour of a  $N^+$ -H to (Piv)C=O hydrogen bond; (4) the (Gly/Ala)N-H gives rise to a two or threecomponent absorption, with one or two minor peaks above  $3400 \text{ cm}^{-1}$  assigned to the free N-H in different environments [39], and a major peak at about 3360  $cm^{-1}$  due to a weakly perturbed N-H bond. The substitution of  $PF_6^-$  for  $BPh_4^-$  with **PrG** has no influence on the C=O absorptions, but results in a single sharp peak at  $3414 \text{ cm}^{-1}$ . indicating that the above absorption at 3360  $\text{cm}^{-1}$ ,

which is not an intrinsic contribution of the BPh<sub>4</sub><sup>-</sup> anion, is due to weak  $N-H\cdots BPh_4^-$  interaction. Thus the N-H bond is mostly oriented outside of the folded molecule in order to interact with the anion, as in the crystal structure of **PTA** (Figure 3), and the two weak contributions for the free N-H bond in **PTA** probably denote the existence of two other minor conformers with the N-H shielded from the BPh<sub>4</sub><sup>-</sup> anion.

Substitution of MeCN for DCM has very little influence on the C=O stretching frequencies (Table 3), indicating that the N<sup>+</sup>-H···O=C hydrogen bond is retained in this weakly aprotic solvent. However, the broad (rGly/rAla)N-H absorption near 3355 cm<sup>-1</sup>, typical of a solvated N-H site, denotes the dissociation of the amide  $\cdots$  anion interaction. In DMSO, disruption of the N<sup>+</sup>-H···O=C hydrogen bond is illustrated by the low N<sup>+</sup>-H frequency, typical of a solvated site, and the high (Piv)C=O frequency assigned to a free vibrator.

The variation with solvent polarity of the proton resonances for the **PrA** C<sup> $\alpha$ </sup>H-C<sub>r</sub>H<sub>2</sub>-N<sup>+</sup>HMe<sub>2</sub> fragment (Table 4) illustrates the contact of the reduced peptide with the BPh<sub>4</sub><sup>-</sup> anion in CDCl<sub>3</sub>. Moreover, the occurrence of one small and one high vicinal coupling constant (Table 4) is in favour of a highly preferred *gauche* conformation for both C<sup> $\alpha$ </sup>H-C<sub>*r*</sub>H<sub>2</sub> (only the value ' $\psi_2$ ' ~ 60° is compatible with the N<sup>+</sup>-H···O=C interaction) and C<sub>*r*</sub>H<sub>2</sub>-N<sup>+</sup>H (' $\omega_2$ ' ~ ± 60°) fragments in CDCl<sub>3</sub> and MeCN-d<sub>3</sub> [34, 36]. The small NH-C<sup> $\alpha$ </sup>H coupling constant (J<sub>MY</sub>) in CDCl<sub>3</sub> is compatible with the folded structure

Compound, solvent	Gly/Ala N-H	rAla/rGly N-H	$N^+-H$	Piv/Boc C=O	Pro/Gly/Ala C=O
<b>PrA</b> , BPh <sub>4</sub> <sup>-</sup>	· · · · · · · · · · · · · · · · · · ·	3431 <sup>w</sup>			
DCM		3416 <sup>w</sup> ; 3360 <sup>s</sup>	2700 <sup>c</sup>	1583	1687
MeCN		3354 <sup>b</sup>	2700 <sup>c</sup>	1582	1685
DMSO		$3260^{\mathrm{b}}$	$2500^{ m bc}$	1617	1679
<b>PrG</b> , BPh <sub>4</sub> <sup>-</sup>					
DCM		3427 <sup>w</sup> ; <i>3359</i> <sup>s</sup>	2700 <sup>c</sup>	1585	1688
MeCN		3359 <sup>b</sup>	$2700^{\circ}$	1583	1688
DMSO		$3265^{\text{b}}$	$2500^{\rm bc}$	1617	1682
<b>PrG</b> , $PF_6^-$					
DCM		3414 <sup>s</sup>	$2700^{\circ}$	1584	1687
<b>GrA</b> , BPh <sub>4</sub> <sup>-</sup>					
DCM	3448 <sup>m</sup> ; 3406 <sup>m</sup>	3365 <sup>s</sup>	3100 <sup>w</sup> ; 2700 <sup>c</sup>	1721 <sup>w</sup> ; <i>1689<sup>m</sup></i>	1678
MeCN	3360 <sup>b</sup>	3360 <sup>b</sup>	2700 <sup>c</sup>	1717 <sup>m</sup> ; <i>1694</i> <sup>w</sup>	1681
ArP, BPh <sub>4</sub> <sup>-</sup>					
DCM	3435 <sup>s</sup>		3000 <sup>c</sup>	1710; 1626	
MeCN	3427 <sup>w</sup> ; 3384 <sup>b</sup>		3000 <sup>c</sup>	1711	1648 <sup>w</sup> ; 1628 <sup>m</sup>
DMSO	$3250^{\mathrm{b}}$		$2500^{\rm bc}$	1702	1650 <sup>s</sup>
<b>rP</b> , PF <sub>6</sub> <sup></sup>					
DCM			3245 <sup>w</sup> ; 3000 <sup>c</sup>	1602 <sup>m</sup> ; 1584 <sup>m</sup>	

Table 3 N–H, N<sup>+</sup>–H and C=O Stretching Frequencies (cm<sup>-1</sup>) for the Protonated Reduced Dipeptides Associated with the BPh<sub>4</sub><sup>-</sup> or PF<sub>6</sub><sup>-</sup> Anion in Various Solvents<sup>a</sup>

<sup>a</sup>Strong (s), medium (m) and weak (w) absorptions. The frequencies in roman and italics denote free and intramolecularly hydrogen bonded vibrators, respectively.

<sup>b</sup>Absorption of a solvated vibrator.

<sup>c</sup>Average frequency for a multicomponent absorption band.

observed in the crystal, with a cisoid disposition  $(\phi' \sim 60^\circ)$  of the NH-C<sup>x</sup>H fragment (Figure 2) [33]. The increase of  $J_{MY}$  in the ionic pair-breaking solvent MeCN-d<sub>3</sub> (Table 4) denotes the appearance of some NH-C<sup>x</sup>H transoid conformer. In DMSO-d<sub>6</sub>, the magnetic equivalence of the C<sub>r</sub>H<sub>2</sub> protons supports a flexible C<sup>x</sup>H-C<sub>r</sub>H<sub>2</sub> fragment, and therefore an open structure of **PrA** in this strong solvating medium.

The folded structures of **PrA** and **PrG** have been investigated by SYBYL molecular dynamics simulation with substitution of an acetyl for the pivaloyl group. The simulation was started from the extended structure for 200 ps at 400 K, generating 200,000 conformers which cover the whole conformational space accessible to these small molecules. After the first 20 ps for stabilization of the temperature at 400 K, one of every 20 conformers set was retained for 0.02 ps intervals, and among the 9000 conformers generated, we have considered those having a short  $(N^+)H\cdots O(Ac)$  distance (<2.5 Å), typical of a  $N^+-H\cdots O=C$  hydrogen bond, and a molecular energy of less than 10 kcal above the

## Table 4 NMR Data for the $NH-C^{\alpha}H-C_{r}H_{2}-NH^{+}$ System in **PrA**

H <sub>Y</sub>	Hм	HA	Hx	
-N-	C <sup>α</sup>	-C,-	N+	Me
	Мe	Н <sub>В</sub>	Ńе	

Chemical shifts (p.p.m.)					Coupling constants (Hz)						
Solvent	$\delta_{\mathbf{A}}$	$\delta_{\mathbf{B}}$	$\delta_{\mathbf{M}}$	$\delta_{\mathbf{X}}$	$\delta_{\mathbf{Y}}$	$J_{AM}$	$J_{BM}$	$J_{AB}$	$J_{AX}$	$J_{\rm BX}$	$J_{MY}$
CDCl <sub>3</sub>	3.21	2.76	3.70	8.05	5.32	11.8	3.5	12.5	1.5	9.5	6.5
MeCN-d <sub>3</sub>	3.22	3.04	4.10	8.04	6.72	11.4	3.9	12.9	1.5	9.3	7.3
DMSO-d <sub>6</sub>	3.	09	4.10	8.82	7.93	6	.7	-	a	a	8.2

<sup>a</sup>Not visible.



Figure 4 rGly/rAla-' $\phi_2$ ', ' $\psi_2$ ' Ramachandran map showing the different conformers folded by an N<sup>+</sup>-H···O=C hydrogen bond, and generated by SYBYL molecular dynamics for Ac-Pro-rGly-N<sup>+</sup>HMe<sub>2</sub> (upper) and Ac-ProrAla-N<sup>+</sup>HMe<sub>2</sub> (lower). The conformations of the rGly and rAla residues in the crystal structures of **PrG** and **PrA** are indicated for comparison.



Figure 5 Four  $\beta$ -like folded conformers of Ac-Pro-rGly-N<sup>+</sup>HMe<sub>2</sub> presenting an N<sup>+</sup>-H···O=C hydrogen bond of the  $i+3 \rightarrow i$  type (SYBYL minimization).

absolute minimum. The Ramachandran maps in Figure 4 show the existence of four folded conformers for **PrG**, and only three for **PrA**, differing in the sign of the rGly/rAla ' $\psi_2$ ' angle, and in the orientation of the Pro-rXaa amide plane. In Table 5 are listed the main conformational data for the energy minimized folded conformers (Figure 5), which are denoted with reference to the cognate  $\beta$ -turn with the sign of the ' $\psi_2$ ' angle indicated in superscript. The sterical hindrances due to the (rAla)C<sup> $\beta$ </sup>H<sub>3</sub> side chain with the Pro oxygen and one of the N<sup>+</sup>Me<sub>2</sub> methyl groups is responsible for the absence of the  $\beta$ II<sup>-</sup> folded structure for **PrA**.

For both **PrG** and **PrA**, the  $\beta$ I<sup>+</sup>-turn is the most frequent folded form according to SYBYL simulation

	Pro		rGly/rAla		H···O	Energy	а
Compound	$\phi$	ψ	'φ'	'ψ'	(Å)	(kcal)	
Ac-Pro-rGly-N <sup>+</sup> HMe <sub>2</sub>							
$\beta$ I <sup>+</sup>	-64	-23	-104	53	1.79	6.2	1250
$\beta I^-$	-66	-20	-66	-40	1.78	6.7	138
$\beta$ II <sup>+</sup>	-65	106	75	45	1.75	8.2	230
$eta$ II $^-$	-66	97	128	-49	1.76	8.5	102
Ac-Pro-rAla-N <sup>+</sup> HMe <sub>2</sub>							
$\beta I^+$	49	-35	~101	57	1.73	5.6	788
$\beta$ I $^-$	-65	-30	-59	-40	1.79	8.2	133
$\beta$ II <sup>+</sup>	-50	131	60	42	1.77	7.4	255

Table 5Conformational Data According to Molecular Simulation for the Folded Structures of Two ReducedPeptides with the Pro-rGly and Pro-rAla Sequences

<sup>a</sup>Number of folded conformers in each family among the 9000 conformers retained after molecular dynamics, and presenting a short  $H \cdots O$  distance (< 2.5 Å) (see Figure 4).

(Figure 4) whereas only  $\beta II^+$  (**PrA** and **PrG**) and  $\beta II^-$ (**PrG**, **PrG**' and **GrG**') are observed in the solid state (Table 2). The simulation was carried out disregarding the peptide  $\cdots$  anion interaction which requires the outer orientation of the amide N—H as in  $\beta II^$ and  $\beta II^+$ . Under these conditions, the inner orientation of the N—H bond as in  $\beta I^+$  (Figure 5) becomes favoured, exactly as in the ion pair dissociating solvent MeCN, where the increase of the NH-C<sup>a</sup>H coupling constant for **PrA** (Table 4) probably reflects the transition from a cisoid ( $\beta II^+$ ) to a transoid ( $\beta I^+$ ) arrangement of the rAla NH-C<sup>a</sup>H system.

#### **GrA Reduced Dipeptide**

The Gly-Ala sequence is not frequently found in  $\beta$ turns [38], and only one-third of the Piv-Gly-Ala-NHMe molecules is  $\beta$ -folded in CHCl<sub>3</sub> [40]. The IR spectrum of GrA in DCM exhibits three C=O and three N-H contributions (Table 3). The weak  $N^+$ -H contribution at 3100  $\text{cm}^{-1}$ , compensated by a broad and multicomponent absorption band about 2700 cm<sup>-1</sup>, and the double absorption of (Boc)C=O at 1721 and 1689  $\text{cm}^{-1}$ , denote a conformational equilibrium between an open conformer (35% occurrence) and a folded conformer (65% occurrence) presenting a  $N^+$ -H···O=C(Boc) hydrogen bond. All the other frequencies are typical of free vibrators, except the absorption at  $3365 \text{ cm}^{-1}$  due to the (rAla)N-H...BPhPro4<sup>-</sup> interaction, also responsible for the shielded (Gly) $C^{\alpha}H_2$  (3.33 p.p.m.) and (rAla) $C^{\alpha}H$  proton (3.71 p.p.m.) resonances in CDCl<sub>3</sub>, compared with 3.64 and 4.27 p.p.m., respectively, in MeCN-d<sub>3</sub>. In MeCN, the relative peak intensities of the (Boc)C=O absorptions (65/35 in favour of the free contribution) are inverted with reference to the CHCl<sub>3</sub>.

#### rP and ArP Reduced Peptides

The Xaa-Pro peptide sequences are not frequently  $\beta$ -folded, except in  $\beta$ VI-turns where the Xaa-Pro amide bond adopts a *cis* disposition [3, 41, 42]. Both **rP** (Piv)C=O and N<sup>+</sup>-H vibrators exhibit two double absorptions in DCM (Table 3), denoting the occurrence of an intramolecular N<sup>+</sup>-H···O=C interaction, closing a seven-membered cycle, in half of the molecules. Under the same conditions, the cognate Piv-Pro-NHiPr molecule exhibits only a very small percentage of the N-H···O=C interaction of the same  $i+2 \rightarrow i$  type [43]. The high (Boc)C=O absorption at 1710 cm<sup>-1</sup> for **ArP** in DCM, typical of a free vibrator, excludes the possibility of a  $\beta$ -like folded



Figure 6  $\gamma$ -like folded conformation of Ac-rPro-N<sup>+</sup>H<sub>2</sub>Me presenting an N<sup>+</sup>-H···O=C hydrogen bond of the  $i+2 \rightarrow i$  type (SYBYL minimization).

structure (Table 3). On the other hand, the low (Ala)C=O and N<sup>+</sup>—H stretching frequencies demonstrate that these sites are hydrogen-bonded in DCM (Table 3). Contrary to the cognate peptide Piv-Ala-Pro-NHMe [44], no *cis* Ala-rPro amide bond is detected for **ArP** in CDCl<sub>3</sub>. It follows that both **rP** and **ArP** adopt in DCM the same  $\gamma$ -like folded structure stabilized by a N<sup>+</sup>—H…O=C hydrogen bond closing a seven-membered cycle [38], which imposes the *trans* disposition on the rPro-preceding amide bond (Figure 6).

This structure, similar to that adopted by the rPro residue in the crystallized neutral reduced tripeptide Boc-rPro-Leu-Gly-NH<sub>2</sub> [21], is essentially retained in MeCN where a small contribution of the free (Ala)C=O carbonyl appears at 1648 cm<sup>-1</sup>, and where 30% of the Ala-rPro amide bonds adopt the *cis* conformation. The  $\gamma$ -like folded structure disappears completely by solvation in DMSO (Table 3).

## **GENERAL CONSIDERATIONS**

In order to investigate the possible influence of a reduced amide bond in a peptide chain, we have considered reduced analogues of peptides known to adopt preferentially the  $\gamma$ - or  $\beta$ -turn structure, where the reduced amide bond has been introduced at the C-terminal position. Owing to the p $K_a$  value of the reduced amide (amine) group, the reduced analogues have been studied in their protonated form associated with a bulky and weakly polar anion in order to minimize the peptide  $\cdots$  anion interactions.

Although the protonated reduced amide group  $C^{\alpha}C_r-N^+-C^{\alpha}$  seems to prefer the transoid conformation, the absence of electronic conjugation cannot exclude the possibility of a *gauche* disposition. Nevertheless, the transoid form exhibits overall dimensions quite similar to those of the peptide

unit. On these bases, it appears to mimic the amide transient state during enzymatic cleavage more closely than the phosphinic  $(PO_2^--CH_2)$  [45], sulphinamide (SO--NH) [46] or sulphonamide  $(SO_2--NH)$  [47] group.

The protonated reduced amide group is a very active proton donor engaged in short contacts with the peptide carbonyls. Hydrogen bonds of the  $i+2 \rightarrow i$  and  $i+3 \rightarrow i$  types, giving rise to  $\gamma$ - and  $\beta$ like folded structures, respectively, have been shown in the present work. The former is especially favoured with the reduced rPro residue, and it appears to be more stable against solvation than the homologous  $\gamma$ -turn in peptides. The latter is also more stable than the  $\beta$ -turn for the cognate peptide sequence, but assumes different conformations differing both in the orientation of the central amide bond and the  $q^+$  or  $q^-$  disposition of the N-C<sup> $\alpha$ </sup>-C<sub>r</sub>-N<sup>+</sup> system. The type of the  $\beta$ -like folded structure, i.e. the orientation of the middle amide bond, not only depends on the sequence, but also on eventual intermolecular interactions such as the peptide ··· anion interaction in the present case.

It is highly probable that the preferential interactions of the ammonium group would depend on the environment, and particularly on the presence and the nature of an anionic counterpart. We have shown that a negative charge distributed on a large surface by electronic conjugation is not tightly bound to the ammonium group. On the contrary, we can envisage that a negative charge located on a limited site, creating a strong local electrical field, would allow stronger ionic interactions than in the present case.

### **EXPERIMENTAL PART**

## **Synthesis**

The reduced dipeptides we have prepared are listed in Table 1 with their abbreviated code derived from the one-letter code. The pivaloyl group has been used to prevent *cis/trans* isomerization of the tertiary Pro-preceding amide bond [26] and the N<sup>+</sup>HMe<sub>2</sub> terminus, having a single N<sup>+</sup>-H bond, to give easily interpretable IR and NMR data. The BPh<sub>4</sub><sup>-</sup> anion has been introduced by anion exchange between the reduced peptide hydrochloride salt and Na<sup>+</sup>BPh<sub>4</sub><sup>-</sup> (Fluka) in water where the reduced peptide BPh<sub>4</sub><sup>-</sup> salt precipitates. The PF<sub>6</sub><sup>-</sup> salt has been obtained by lyophilizing a water solution of the neutral reduced peptide and hexafluorophosphoric acid diethyloxide (Aldrich). The neutral reduced peptides are stable, but often obtained as an oily, more or less carbonated product. The  $PF_6^-$  and  $BPh_4^-$  reduced peptide salts are more stable in the solid state than in solution where the rapid evolution with time of their IR and NMR data denotes a chemical instability preventing long-term NMR experiments.

The syntheses of **PrG**, **PrG**' and **GrG**' have been already reported [22, 23]. **GrA** has been prepared from Boc-Gly-ONp and the racemic NH<sub>2</sub>-CHMe-CH<sub>2</sub>-NMe<sub>2</sub> diamine (Fluka) abbreviated as H-DL-rAla-NMe<sub>2</sub>. Coupling of Piv-Pro-OH and H-DL-rAla-NMe<sub>2</sub>, using the mixed anhydride with isobutyl chloroformate, gave a mixture of the two Piv-L-Pro-L-rAla-NMe<sub>2</sub> and Piv-L-Pro-D-rAla-NMe<sub>2</sub> diastereoisomers. The former (**PrA**) eluted first by flashchromatography with ethanol on silica gel, and its stereochemistry was validated by X-ray crystallography (see below).

**Boc-Giy-rAla-NMe<sub>2</sub>.** Racemic oil,  $R_{\rm F} = 0.12$  (EtOH). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.18 (d, rAla- $C^{\beta}H_{3}$ , J = 6.5 Hz); 1.45 (s, Boc-( $CH_{3}$ )<sub>3</sub>); 2.21 (s, N<sup>+</sup>( $CH_{3}$ )<sub>2</sub>); 2.35 (A) and 2.18 (B) (ABX, rAla- $C_{r}H_{2}$ ,  $J_{AB} = 12.4$  Hz,  $J_{AX} = 9.4$  Hz,  $J_{BX} = 5.5$  Hz); 3.77 (d, Gly- $C^{\alpha}H_{2}$ , J = 5.7 Hz); 3.97 (m, rAla- $C^{\alpha}H$ ); 5.32 (d, rAla-NH); 6.38 (t, Gly-NH). IR (film): 1710 cm<sup>-1</sup> (Boc-*CO*); 1645 cm<sup>-1</sup> (Gly-*CO*).

**Boc-Giy-rAla-N<sup>+</sup>HMe<sub>2</sub>, BPh<sub>4</sub><sup>-</sup> (GrA).** Racemic, <sup>1</sup>H-NMR (MeCN-d<sub>3</sub>): 1.18 (d, rAla- $C^{\beta}H_{3}$ , J=6.8 Hz); 1.42 (s, Boc-( $CH_{3}$ )<sub>3</sub>); 2.85 (s, N<sup>+</sup>( $CH_{3}$ )<sub>2</sub>); 3.07 (d, rAla- $C_{r}H_{2}$ , J=6.7 Hz); 3.64 (d, Gly- $C^{\alpha}H_{2}$ , J=5.7 Hz); 4.27 (m, rAla- $C^{\alpha}H$ ); 5.72 (b, Gly-NH); 6.80–7.32 (m, B( $C_{6}H_{5}$ )<sub>4</sub> + rAla-NH + N<sup>+</sup>H). IR (MeCN): 1681 cm<sup>-1</sup> (Ala-CO); 1717 cm<sup>-1</sup> (Boc-CO); 2700 cm<sup>-1</sup> (N<sup>+</sup>H); 3360 cm<sup>-1</sup> (Gly-NH + rAla-NH).

**Piv-Pro-rAla-NMe**<sub>2</sub>. Melting point m.p. = 82°C,  $R_{\rm F} = 0.23$  (EtOH) and 0.36 (MeOH/DCM 40/60 v/v). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.14 (d, rAla-C<sup> $\beta$ </sup><u>H</u><sub>3</sub>, J = 6.5 Hz); 1.27 (s, Boc-(C<u>H</u><sub>3</sub>)<sub>3</sub>; 1.80–2.26 (m, Pro-C<sup> $\beta$ </sup><u>H</u><sub>2</sub> + C<sup> $\gamma$ </sup><u>H</u><sub>2</sub>); 2.22 (s, N<sup>+</sup>(C<u>H</u><sub>3</sub>)<sub>2</sub>); 2.36 (A) and 2.16 (B) (ABX, rAla-C<sub>T</sub><u>H</u><sub>2</sub>,  $J_{\rm AB} = 12.2$  Hz,  $J_{\rm AX} = 5.9$  Hz,  $J_{\rm BX} = 8.9$  Hz); 3.70 (m, Pro-C<sup> $\delta$ </sup><u>H</u><sub>2</sub>); 3.91 (m, rAla-C<sup> $\alpha$ </sup>H); 4.58 (m, Pro-C<sup> $\alpha$ </sup><u>H</u>); 6.69 (d, rAla-N<u>H</u>, J = 4.4 Hz). IR (KBr): 1625 cm<sup>-1</sup> (Piv-CO); 1665 cm<sup>-1</sup> (Pro-CO).

**Piv-Pro-rAla-N<sup>+</sup>HMe<sub>2</sub>,BPh<sub>4</sub><sup>-</sup>** (PrA).  $[\alpha]_D^{20} = -10.8$ (c=1, MeOH), <sup>1</sup>H-NMR (MeCN-d<sub>3</sub>): 1.21 (d, rAla-C<sup> $\beta$ </sup>H<sub>3</sub>, J=7.1 Hz); 1.25 (s, Boc-(CH<sub>3</sub>)<sub>3</sub>); 1.70-2.32



Figure 7 Synthesis of **ArP** by the nucleophilic substitution procedure.

(m, Pro- $C^{\beta}\underline{H}_{2} + C^{\gamma}\underline{H}_{2}$ ); 2.87 (s, N<sup>+</sup>( $C\underline{H}_{3}$ )<sub>2</sub>); 3.22 (A), 3.04 (B) (ABX, rAla- $C_{\Gamma}\underline{H}_{2}$ ,  $J_{AB} = 12.9$  Hz,  $J_{AX} = 3.8$  Hz,  $J_{BX} = 11.8$  Hz); 3.75 (m, Pro- $C^{\delta}\underline{H}_{2}$ ); 4.00–4.35 (m, Pro- $C^{\alpha}\underline{H}$  + rAla- $C^{\alpha}\underline{H}$ ); 6.72 (d, rAla-N $\underline{H}$ , J = 7.3 Hz); 6.75–7.20 (m, B( $C_{6}\underline{H}_{5}$ )<sub>4</sub>); 8.04 (b, N<sup>+</sup> $\underline{H}$ ). IR (MeCN): 1617 cm<sup>-1</sup> (Piv-*CO*); 1679 cm<sup>-1</sup> (Pro-*CO*); 2700 cm<sup>-1</sup> (N<sup>+</sup> $\underline{H}$ ); 3354 cm<sup>-1</sup> (rAla- $N\underline{H}$ ).

**rP** was prepared by reductive animation of the racemic Boc-prolinal [27], and **ArP** by nucleophilic substitution (Figure 7) of Boc-rPro-Br, obtained by bromination of Boc-prolinol (Merck) [28]. In both cases, the temporary Z-protection of the reduced amide function was required to prevent further undesirable acylation.

**Piv-rPro-NHiPr.** Racemic oil.  $R_{\rm F} = 0.20$  (iPrOH/DCM 10/90 v/v). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.32 (m, Piv-(C<u>H</u><sub>3</sub>)<sub>3</sub> + iPr-(C<u>H</u><sub>3</sub>)<sub>2</sub>); 1.78 (m, rPro-C<sup> $\beta$ </sup><u>H</u>); 1.92–2.09 (m, Pro-C<sup> $\gamma$ </sup><u>H</u><sub>2</sub>); 2.22 (m, rPro-C<sup> $\beta$ </sup><u>H</u>); 3.15 (A) and 2.93 (B) (ABX, rPro-C<sub>r</sub><u>H</u><sub>2</sub>,  $J_{\rm AB} = 12.4$  Hz,  $J_{\rm AX} = 3.5$  Hz,  $J_{\rm BX} = 6.8$  Hz); 3.18 (m, iPr-C<u>H</u>); 3.60 and 3.87 (m, rPro-C<sup> $\delta$ </sup><u>H</u><sub>2</sub>); 4.36 (m, rPro-C<sup> $\alpha$ </sup><u>H</u>); 7.66 (d, iPr-N<u>H</u>)). IR (film): 1625 cm<sup>-1</sup> (Piv-CO).

**Piv-rPro-N<sup>+</sup>H<sub>2</sub>iPr**, **PF**<sub>6</sub><sup>-</sup> (**rP**). Racemic. <sup>1</sup>H-NMR (CDCl<sub>3</sub>/DMSO-d<sub>6</sub> 90/10): 0.94 (s, Piv-(C<u>H</u><sub>3</sub>)<sub>3</sub>); 0.98 and 1.00 (2d, iPr-(C<u>H</u><sub>3</sub>)<sub>2</sub>, J = 6.4 Hz); 1.25–1.92(m, rPro-C<sup> $\beta$ </sup><u>H</u><sub>2</sub> + C<sup> $\gamma$ </sup><u>H</u><sub>2</sub>); 2.70 (m, rPro-C<sub>r</sub><u>H</u><sub>2</sub>); 3.26 and 3.51 (m, rPro-C<sup> $\delta$ </sup><u>H</u><sub>2</sub>); 3.72 (m, iPro-C<u>H</u>); 3.85 (m, rPro-C<sup> $\alpha$ </sup><u>H</u>); 8.46 and 8.76 (b, N<sup>+</sup><u>H</u><sub>2</sub>). IR (KBr): 1595 cm<sup>-1</sup> (Piv-CO); 2800 cm<sup>-1</sup> (N<sup>+</sup>H<sub>2</sub>).

**Boc-Ala-rPro-NHMe.** Oil.  $R_{\rm F} = 0.32$  (iPrOH/DCM 7/93 v/v). <sup>1</sup>H-NMR (CDCl<sub>3</sub>): 1.35 (d, Ala-C<sup> $\beta$ </sup> $\underline{H}_{3}$ , J = 7.0 Hz); 1.41 (s, Boc-(C $\underline{H}_{3}$ )<sub>3</sub>); 1.60–2.40 (m, rPro-C<sup> $\beta$ </sup> $\underline{H}_{2}$  + rPro-C<sup> $\gamma$ </sup> $\underline{H}_{2}$  + Me-N $\underline{H}$ ); 2.73 (s, N-C $\underline{H}_{3}$ ); 2.92–3.21 (m, rPro-C<sub>r</sub> $\underline{H}_{2}$ ); 3.55 and 3.81 (m, rPro-

 $C^{\delta}\underline{H}_{2}$ ; 4.46 (m, rPro- $C^{\alpha}\underline{H}$ ) + Ala- $C^{\alpha}\underline{H}$ ); 5.56 (d, Ala-N<u>H</u>), J=7.9 Hz. IR (film): 1710 cm<sup>-1</sup> (Boc-*CO*); 1645 cm<sup>-1</sup> (Ala-*CO*).

**Boc-Ala-***i***Pro-***N*<sup>+</sup>*H*<sub>2</sub>*Me*, *BPh*<sub>4</sub><sup>-</sup> (**ArP**).  $[\alpha]_D^{20} = -32.6$ (*c* = 1, MeOH), <sup>1</sup>H-NMR (MeCN-d<sub>3</sub>): *trans* (t)/*cis* (c) 70/30; 1.24 (c) and 1.26 (t) (d, Ala-C<sup> $\beta$ </sup><u>H</u><sub>3</sub>, *J* = 6.8 Hz); 1.38 (t) and 1.44 (c) (s, Boc-(C<u>H</u><sub>3</sub>)<sub>3</sub>); 1.60–2.38 (m, rPro-C<sup> $\beta$ </sup><u>H</u><sub>2</sub> + C<sup> $\gamma$ </sup><u>H</u><sub>2</sub>); 2.61 (c) and 2.63 (t) (s, N<sup>+</sup>-C<u>H</u><sub>3</sub>); 3.04 (m, rPro-C<sub>r</sub><u>H</u><sub>2</sub>); 3.47 and 3.75 (m, rPro-C<sup> $\delta$ </sup><u>H</u><sub>2</sub>); 4.13 (m, rPro-C<sup> $\alpha$ </sup><u>H</u>); 4.37 (m, Ala-C<sup> $\alpha$ </sup><u>H</u>); 5.58 (t) and 5.81 (c) (b, Ala-N<u>H</u>); 6.71–7.80 (m, B(C<sub>6</sub><u>H</u><sub>5</sub>)<sub>4</sub> + N<sup>+</sup><u>H</u><sub>2</sub>).

#### X-ray Diffraction

Single crystals of PrA were grown by cooling an AcOEt solution (orthorhombic,  $P2_12_12_1$ , a=11.322(1) Å, b = 14.222(2) Å, c = 22.002(3) Å, Z = 4, calculated density  $1.15 \text{ g/cm}^3$ ). The X-ray diffraction data were collected on an Enraf Nonius CAD-4 four-circle diffractometer in the  $\omega/2\theta$ -scan mode. Reflections numbering 3788 in total, of which 2212 were observed with  $I > \sigma(I)$ , were measured at room temperature in the 1-70°  $\theta$  range using Cu-Ka radiation ( $\lambda = 1.54178$  Å) monochromatized by a graphite crystal. During data collection, two standard reflections were measured every 2 h to check the stability of the crystal. Intensities were corrected for Lorentz and polarization effects but no absorption correction was applied. The crystal structure was solved by direct methods using SHELXS 90 [29]. The E-maps revealed the whole molecule except the hydrogen atoms, and the structure was refined through the least-squares procedure with the complete matrix of normal equations [30]. Heavy atoms were affected by anisotropic thermal factors, and hydrogen atoms were located on E-map differences and affected by an isotropic thermal factor of  $4 \text{ Å}^2$ . The residual R factors are R = 0.056 and  $R_w = 0.057$  $(w = 0.056 / [\sigma^2(F) + 0.00084 F^2])$ , with largest difference peak and hole of 0.19 and  $-0.16 \text{ e/Å}^3$ , respectively. The NH hydrogen atoms were placed 1.03 Å from N in the direction obtained by refinement [31].

## FT-IR and <sup>1</sup>H-NMR Spectroscopy

IR spectra were run in the Fourier transform mode on a Bruker IFS-25 apparatus using a cell path of 0.5 mm in order to investigate the N—H (3200– 3500 cm<sup>-1</sup>), C=O (1580–1720 cm<sup>-1</sup>) and N<sup>+</sup>—H (2500–3300 cm<sup>-1</sup>) stretching frequencies in DCM, MeCN and DMSO. The peptide concentration was 0.005 M, and further dilution confirmed the absence of any molecular aggregation. The N-H and C=O stretching frequencies were assigned on the basis of previous studies on peptides [32] and ammonium groups [24, 25]. The free amide N-H is expected to give a sharp absorption at  $3400-3450 \text{ cm}^{-1}$ , and the free (Piv)C=O at  $1620-1625 \text{ cm}^{-1}$  in DCM. When engaged in an intramolecular hydrogen bond, their absorption is shifted to lower frequencies by  $100-200 \text{ cm}^{-1}$  and  $10-25 \text{ cm}^{-1}$ , respectively. The  $N^+H$  and  $N^+H_2$  stretching frequencies greatly depend both on the solvent and on the associated anion. In a weakly polar solvent such as DCM, the N<sup>+</sup>H bond gives rise to a sharp peak at about 3250 or  $3100 \text{ cm}^{-1}$  when associated with the PF<sub>6</sub><sup>-</sup> or BPh<sub>4</sub><sup>-</sup> anion, respectively, which turns into a multicomponent and broad absorption shifted down to 2500-3000  $\text{cm}^{-1}$  upon formation of an  $N^+\text{--}H\cdots\text{O}\text{=}C$ hydrogen bond in DMA [25]. The same is true for the  $N^+H_2$  cation which absorbs at 3245 or 3140 cm<sup>-1</sup> in DCM when associated with the  $PF_6^$ or  $BPh_4^-$  anion, respectively. The  $N^+H_2$  cation also presents a very weak absorption at 1596  $cm^{-1}$  in DCM. Both the  $PF_6^-$  or  $BPh_4^-$  anions have no visible absorption in the 2200-2800 and 3150- $3600 \text{ cm}^{-1}$  regions, but  $BPh_4^-$  exhibits a weak and sharp aromatic contribution at 1580  $\text{cm}^{-1}$ .

<sup>1</sup>H-NMR spectra were run on a Bruker AC-200P apparatus with Me<sub>4</sub>Si as internal reference, and spin systems have been solved by COSY and TOCSY experiments. The vicinal coupling constants *J* in the NH-C<sup> $\alpha$ </sup>H-C<sub>r</sub>H<sub>2</sub>-N<sup>+</sup>H moiety have been exploited in terms of torsional angles from the Karplus correlations adapted to the NH-C<sup> $\alpha$ </sup>H [33] and C<sup> $\alpha$ </sup>H-C<sup> $\beta$ </sup>H<sub>2</sub> fragments [34], provided the electronegativity correction *J* (corrected) = 1.06 *J* (observed) is applied [35], and to the CH<sub>2</sub>-N<sup>+</sup>H fragment [36].

#### Acknowledgements

The authors thank D. Mangeot and J. M. Grosse for technical assistance.

## REFERENCES

- M. Szelke, B. J. Leckie, M. Tree, A. Brown, J. Grant, A. Hallett, M. Hughes, D. M. Jones and A. F. Lever (1982). H-77: A potent new renin inhibitor. *In vitro* and *in vivo* studies. *Hypertension*. *Supp. II* 4, 52–69.
- J. Cooper, S. Foundling, A. Hemming, T. Blundell, D. M. Jones, A. Hallett and M. Szelke (1987). The structure of a synthetic pepsin inhibitor complexed with endothiapepsin. *Eur. J. Biochem.* 169, 215–221.

- D. E. Epps, B. Mao, D. J. Staples and T. K. Sawyer (1988). Structure-conformation relationships of synthetic peptide inhibitors of human renin studied by resonance energy transfer and molecular modeling. *Int. J. Peptide Protein Res.* 31, 22–34.
- T. K. Sawyer, D. T. Pals, B. Mao, L. L. Maggiora, D. J. Staples, A. E. De Vaux, H. J. Shostarez, J. H. Kinner and C. W. Smith (1988). Structure-conformationactivity relationships of renin inhibitory peptides having P<sub>1</sub>-P'<sub>1</sub> Xaaµ[CH<sub>2</sub>NH]Yaa substitutions: molecular modeling and crystallographic studies. *Tetrahedron* 44, 661-673.
- 5. T. K. Sawyer, D. T. Pals, B. Mao, D. J. Staples, A. E. De Vaux, L. L. Maggiora, J. A. Affholter, W. Kati, D. Duchamp, J. B. Hester, C. W. Smith, H. H. Saneii, J. Kinner, M. Handschumacher and W. Carlson (1988). Design, structure, activity and molecular modeling studies of potent renin inhibitory peptides having Nterminal N<sup>im</sup>-For-Trp(Ftr): angiotensinogen congeners modified by  $P_1-P'_1$  Phe-Phe, Sta, Leu $\psi$ [CH(OH)CH<sub>2</sub>]Val or Leu $\psi$ [CH<sub>2</sub>NH]Val substitutions. J. Med. Chem. 31, 18–30.
- J. Martinez, J. P. Bali, M. Rodriguez, B. Castro, R. Magous, J. Laur and M. F. Lignon (1985). Synthesis and biological activities of some pseudopeptide analogues of tetragastrin: The importance of the peptide backbone. J. Med. Chem. 28, 1874–1879.
- J. S. Kaltenbronn, J. P. Hudspeth, E. A. Lunney, B. M. Michniewicz, E. D. Nicolaides, J. T. Repine, W. H. Roark, M. A. Stier, F. J. Tinner, P. K. W. Woo and A. D. Essenburg (1990). Renin inhibitors containing isosteric replacements of the amide bond connecting the P<sub>1</sub> and P<sub>2</sub> sites. J. Med. Chem. 33, 838–845.
- J. Martinez, M. Rodriguez, J. P. Bali and J. Laur (1986). Phenethyl ester derivatives analogues of the C-terminal tetrapeptide of gastrin as potent gastrin antagonists. J. Med. Chem. 29, 2201–2206.
- M. Lebl, E. E. Sugg, G. Van Binst, P. Van der Elst, D. Tourwé, J. Slaninova and V. J. Hruby (1987). Analogs of oxytocin containing a modified peptide bond. *Int. J. Peptide Protein Res.* 30, 318–322.
- D. H. Coy, P. Heinz-Erian, N. Y. Jiang, Y. Saaki, J. Taylor, J. P. Moreau, W. T. Wolfrey, J. D. Gardner and R. T. Jensen (1988). Probing peptide backbone in bombesin. A reduced peptide bond analog with potent and specific antagonist activity. J. Biol. Chem. 263, 5056–5060.
- 11. C. Di Bello, A. Scatturin, G. Vertuani, G. D'Auria, M. Gargiulo, L. Paolillo, G. Trivellone, L. Gozzini and R. De Castiglione (1991). Conformational studies of bombesin antagonists: CD and NMR characterization of [Thr<sup>6</sup>,  $Leu^{13}\psi$ [CH<sub>2</sub>NH]Met<sup>14</sup>] bombesin 6–14. *Biopolymers 31*, 1397–1408.
- W. M. Kazmierski, R. D. Ferguson, R. J. Knapp, G. K. Lui, H. I. Yamamura and V. J. Hruby (1992). Reduced peptide bond cyclic somatostatin based opioid octapeptides. Synthesis, conformational properties and

pharmacological characterization. Int. J. Peptide Protein Res. 39, 401-414.

- 13. J. Couder, D. Tourwé, G. Van Binst, J. Schuurkens and J. E. Leysen (1993). Synthesis and biological activities of  $\psi$ [CH<sub>2</sub>-NH] pseudopeptide analogues of the C-terminal hexapeptide of neurotensin. Int. J. Peptide Protein Res. 41, 181-184.
- M. Rodriguez, P. Fulcrand, M. F. Lignon, M. C. Galas, J. P. Bali, R. Magous, P. Dubreuil, J. Laur and J. Martinez in: *Peptides: Chemistry and Biology*, G. R. Marshall, Ed., p. 101-104, ESCOM, Leiden, The Netherlands 1988.
- 15. A. Aumelas, M. Rodriguez, A. Heitz, B. Castro and J. Martinez (1987). <sup>1</sup>H and <sup>13</sup>C NMR studies of pseudopeptide analogues of the C-terminal tetrapeptide of gastrin. *Int. J. Peptide Protein Res.* 30, 596–604.
- P. Van der Elst, M. Elseviers, E. De Cock, M. Van Marsenille, D. Tourwé and G. Van Binst (1986). Synthesis and conformational study of two L-prolyl-Lleucyl-glycinamide analogs with a reduced peptide bond. Int. J. Peptide Protein Res. 27, 633–642.
- S. Ma and A. F. Spatola in: *Peptides, Chemistry and Biology*, J. A. Smith and J. E. Rivier, Eds., p. 777–778, ESCOM, Leiden, The Netherlands 1991.
- 18. S. Ma and A. F. Spatola (1993). Conformations of  $\psi$ [CH<sub>2</sub>-NH] pseudopeptides Cyclo[Gly-Pro $\psi$ [CH<sub>2</sub>-NH]Gly-D-Phe-Pro].TFA and cyclo[Gly-Pro $\psi$ [CH<sub>2</sub>-NH]Gly-D-Phe-Pro]. Int. J. Peptide Protein Res. 41, 204-206.
- P. Dauber-Osguthorpe, D. K. Jones, M. M. Campbell and D. J. Osguthorpe (1990). Reduced and retro reduced peptide analogs. Conformations and energies. *Tetrahedron Lett.* 31, 917–920.
- P. Dauber-Osguthorpe, M. M. Campbell and D. J. Osguthorpe (1991). Conformational analysis of peptide surrogates. Reduced and retro-amide links in blocked alanine and in secondary structure. *Int. J. Peptide Protein Res.* 38, 357–377.
- C. Toniolo, G. Valle, M. Crisma, J. S. Kaltenbronn, J. T. Repine, G. Van Binst, M. Elseviers and D. Tourwé (1989). ψ[CH<sub>2</sub>-NH] backbone-modified peptides: first unequivocal observation of a C<sub>7</sub> structure in a linear peptide. *Peptide Res*, 2, 332-337.
- 22. L. El Masdouri, A. Aubry, C. Sakarellos, E. J. Gomez, M. T. Cung and M. Marraud (1988). A β-turn-like conformation in reduced peptides. Crystal structures ot two dipeptide analogs with Pro-Gly/[CH<sub>2</sub>-NH] sequence. Int. J. Peptide Protein Res. 31, 420-428.
- 23. L. El Masdouri, A. Aubry, V. Grand and M. Marraud (1992). Structure of tBuCO-Gly-Gly $\psi$ [CH<sub>2</sub>-N<sup>+</sup>H<sub>2</sub>]N HEt, BPh<sub>4</sub><sup>-</sup>. Acta Crystallogr. Ser. C 48, 173-175.
- 24. E. Moreno-Gonzalez, M. Marraud and J. Néel (1977). Etude des solutions de sels de trialcoylammonium par spectroscopie infrarouge; interactions entre cations trialcoylammonium et espèces nucléophiles. J. Chim. Phys. 74, 563–571.
- 25. E. Moreno-Gonzalez and M. Marraud (1980). Cation

ammonium tertiaire et liaison hydrogène intramoléculaire. J. Chim. Phys. 77, 149–155.

- H. Nishihara, K. Nishihara, T. Uefuji and N. Sakota (1975). Conformation and circular dichroism of several *N*-acyl-L-prolines. *Bull. Chem. Soc. Jpn* 48, 553–555.
- M. Szelke, B. Leckie, A. Hallet, D. M. Jones, J. Sueiras, B. Atrash and A. F. Lever (1982). Potent new inhibitors of human renin. *Nature 299*, 555–557.
- E. H. Axelrod, G. M. Milne and E. E. Van Tamelen (1970). General 1,5-diene synthesis involving overall alkyl alcohol coupling with geometrical and positional control. J. Am. Chem. Soc. 92, 2139–2141.
- G. M. Sheldrick (1990). Phase annealing in SHELX-90: direct method for large structures. *Acta Crystallogr. Ser.* A. 46, 467–473.
- G. M. Sheldrick: Programs for Crystal Structure Determination. University of Cambridge, Cambridge, UK 1976.
- 31. R. Taylor and O. Kennard (1983). Comparison of X-ray and neutron diffraction results for the N−H…O=C hydrogen bonds. *Acta Crystallogr. Ser. B* 39, 133–138.
- 32. A. Aubry, M. T. Cung and M. Marraud (1985). βI- and βII-turn conformations in model dipeptides with the Pro-Xaa sequences. J. Am. Chem. Soc. 107, 7640– 7647.
- 33. M. T. Cung, M. Marraud and J. Néel (1974). Experimental calibration of a Karplus relationship in order to study the conformations of peptides by nuclear magnetic resonance. *Macromolecules* 7, 606–613.
- M. T. Cung and M. Marraud (1982). Conformational dependence of the vicinal proton coupling constant for the C<sup>α</sup>-C<sup>β</sup> bond in peptides. *Biopolymers* 21, 953–967.
- V. F. Bystrov (1976). Spin-spin coupling and the conformational states of peptide systems. *Prog. Nucl. Res.* 10, 41-81.
- R. R. Fraser, R. N. Renaud, J. K. Saunders and Y. Y. Wigfield (1973). Dihedral angular dependence of H-N-C-H coupling constants. Protonated amines in trifluoroacetic acid. *Can. J. Chem.* 51 2433-2437.
- E. Benedetti in: Chemistry and Biochemistry of Amino Acids, Peptides and Proteins. Vol. 6, B. Weinstein, Ed., p. 105–184, Marcel Dekker Inc., New York, USA 1982.
- G. D. Rose, L. M. Gierasch and J. A. Smith (1985). Turns in peptides and proteins. *Adv. Protein Chem.* 37, 1–109.
- M. Marraud, J. Néel, M. Avignon and V. H. Pham (1970). Contribution à l'étude conformationnelle des composés dipeptidiques en solution. J. Chim. Phys. 67, 959–964.
- 40. G. Boussard and M. Marraud (1985). β-turns in model dipeptides. An infrared quantitative analysis with NMR correlation. J. Am. Chem. Soc. 107, 1825–1828.
- 41. A. Aubry, B. Vitoux and M. Marraud (1985). *N*-méthyl peptides. VIII. Etude radiocristallographique du repliement  $\beta$ VI des séquences homochirales. *J. Chim. Phys.* 82, 933–939.
- 42. M. W. MacArthur and J. M. Thornton (1991). Influence

of proline residues on protein conformation. J. Mol. Biol. 218, 397-412.

- 43. G. Boussard, M. Marraud and A. Aubry (1979). Experimental investigations on the backbond folding of proline-containing model tripeptides. *Biopolymers* 18, 1297-1331.
- 44. B. Vitoux, A. Aubry, M. T. Cung and M. Marraud (1986). *N*-Methyl peptides VII. Conformational perturbations induced by *N*-methylation of model dipeptides. *Int. J. Peptide Protein Res.* 27, 617–632.
- W. H. Parson, A. A. Patchett, H. G. Bull, W. R. Schoen, D. Taub, J. Davidson, P. L. Combs, J. P. Springer, H. Gadebusch, B. Weissberger, M. E. Valiant, T. N. Mellin

and R. D. Busch (1988). Phosphinic acid inhibitors of D-alanyl-D-alanine ligase. J. Med. Chem. 31, 1772-1778.

- D. Merricks, P. G. Sammes, E. R. H. Walker, K. Henrick and M. McPartlin (1991). Some studies on peptide analogues involving the sulphinamide group. *J. Chem.* Soc. Perkin Trans. 1, 2169–2176.
- A. Calcagni, E. Gavuzzo, G. Lucente, F. Mazza, G. Pochetti and D. Rossi (1989). Structure and conformation of peptides containing the sulphonamide junction. I. Synthesis and conformation of cyclo(MeTau-Phe-D-Pro). Int. J. Peptide Protein Res. 34, 319–324.